SYNTHESIS OF TWO LINEAR OCTAPEPTIDE FRAGMENTS OF CYCLOSPORIN BY STRPWISK AND FRACHIONT CONDIDISATION STRATEGIES

I.J.GALPIN^{*}, A.K.A. MOHAMMED, A. PATEL and G. PRIESTLEY

The Robert Robinson Laboratories, The University of Liverpool, P.O.Box 147, Liverpool, L69 3BX, England

(Received in UK 21 December 1987)

Abstract: The Octapeptide Z-(Me)Leu-Val-(Me)Leu-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu was prepared by stepwise elongation starting from H-(Me)Val- $\frac{1}{2}$. Observe the consequence of the consequ throughout as they gave high yields of optically homogeneous products in this extensively <u>N</u>-methylated peptide. The above peptide and the octapeptide Z-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-(Me)Thr(Bu)-Abu-Sar-OBu were also prepared by the fragment consensation approach employing a variety of coupling methods. Ultimately, it was clear that the stepvise assembly gave the highest yield, and most homogeneous product.

The cyclosporins are a group of potent immunosuppressive compounds which have been isolated from <u>Tolypocladium inflatum</u> Gams, $^{\text{1}}$ the general structure of the cyclosporins, based on X-ray crystallographic² and nmr data³ being shown in Figure 1.

Cyclosporin A contains seven relatively hydrophobic, N-methylated amino acids, including N-methyl leucine and N-methyl valine. In addition, all cyclosporins contain the unusual "C_o" amino acid (4R)-4-[(E)-2- butanyl]-4,N-dimethyl-L-threonine) (MeBmt); this residue has the same optical configuration as threonine and has been synthesised in stereochemically pure form. $4-6$ The majority of the other residues are invariant; however, a number of variations are known at position 2 and this gives rise to the range of known cyclosporins.⁷

As the immunosuppressive property of cyclosporin A is known 8.9 to be intimately connected with the nature of the residues at positions one and two, we chose to attempt modification of the cyclosporin structure at these positions. An additional impetus for modification at position-2 was the knowledge that the norvaline-2 analogue of cyclosporin is considerably less nephrotoxic than cyclosporin A itself, which has an aminobutyric acid residue at this position 10 .

The total synthesis of cyclosporins has been reported by two groups, $9,11,12$ and in both, the same general route using the fragment condensation approach with final cyclisation between the alanine residues has been used. These syntheses were facilitated using t-butyloxycarbonyl for amino group protection and a benzyl ester at the C-terminus. In a recent subsequent paper 13 the merits of benzyloxycarbonyl and 9-fluorenylmethoxycarbonyl have been contrasted with the use of t-butyloxycarbonyl for amino group protection. These groups in combination with the $bis(-2-$ oxo-3-oxazolidinyl)phosphinic chloride^{13,14} coupling reagent provided a means of

All amino acids are of the L-configuration unless otherwise specified. and nomenclature follows IUPAC-IUB Joint commission on biochemical nomenclature (JCBN) Nomenclature and symbolism for amino acids and peptides 1983.

assembling fragments of cyclosporins which was comparable to our ${\tt own.}^{15}$

In the current paper we describe the synthesis of the octapeptides (1) and (2) using both the fragment coupling and stepwise elongation approach.

Z-(ne)Leu-Val-(Me)Le"-Ala-D-Ala-(Me)Lau-(Me)Le"-(Me)Val-OB~t (1) Z-Ala-D-Ala-(ne)Leu-(Me)Lau-(Me)Val-(ne)Thr(But)-Abu-Sar-OBut (2)

The synthesis of such peptides in optically pure form is not straightforward as the factors which influence racemisation are complex, $^{15-17}$ and, there is an increased tendency with N </u> methylated amino acids towards racemisation. Recently we have demonstrated¹⁵ that the diphenylphosphinic (Dpp) mixed anhydrides are advantageous for the coupling of N-methyl amino acids when benxyloxycarbonyl is used for amino group protection. This protecting group was generally removed by catalytic hydrogenolysis using palladium on charcoal catalyst. In the initial work, the use of phenyl esters for carboxyl group protection was explored, but it was rapidly apparent that many side reactions occurred in the synthesis of N-methylated peptides using this protection, particularly the formation of diketopiperazine with the concomitant expulsion of phenoxide. In the final strategy the tert-butyl ester protecting group of Anderson¹⁸ and Roeske¹⁹ was used; also, <u>tert</u>-butyl ether protection was used for the side chain hydroxyl group of threonine and hydroxyproline derivatives, as it is widely believed that better yields may be obtained when side chain hydroxyl groups are blocked. The tert-butyl ether and ester groups remain unaffected during peptide bond formation and are easily removed by acidolysis at the appropriate point in the synthesis without the use of basic conditions which might give rise to racemisation.

N-methyl leucine, N-methyl valine and N-methyl threonine which were required for this work were prepared by the N-methylation procedure reported by Benoiton^{20,21}. . The N-methylation of leucine and valine was straightforward; however, difficulties, were encountered with the synthesis of N-methyl threonine. Mass spectrometry of the product of N-methylation of Z-Thr(OBu^t)-OH showed two intense peaks at m/z 324 and 310, corresponding to the methylated and **non-methylatad threonine derivative respectively. The proton n.m.r. spectrum of tha product showed a broad signal at 6 5.8 due to the NH resonance,this being in addition to the signal at 6** 3.1 which was due to the N-methyl group protons. Thus only partial N-methylation had occured **in line with previous observations by McDermott and Benoiton. 20 This difficulty may be attributed to staric hindrance of the side chain, as on attempted re-methylation of the partially methylated compound no change in the spectroscopic data was observed. Purified** Z-(Me)Thr(Bu^t)-OH was finally obtained by separating the two compounds by silica gel chromato**graphy.**

The octapeptides (1) and (2) were initially synthesised by the fragment condensation approach, octapeptide (1) being simultaneously synthesised using a stepvise procedure. The usa of the fragment condensation approach was investigated, as it would reduce the length of time taken for the synthesis since the two subfragments could be prepared in parallel; also the fragment condensation approach had been used successfully by other workers. 9.11.12 The protected tripeptides (4) and (6) and the protected pentapeptide (10) were prepared by stepwise build-up according to the routes shown in Schemes 1 - 3. A variety of reaction conditions were studied, l5 and it was ultimately found that the diphenylphosphinic mixed anhydride procedure gave superior results in terms of both yield and optical homogeneity.

SCHEME 1 Synthesis of the protected (4-6) fragment.

SCHEME 2 Synthesis of the protected (1-3) fragment

In order to carry out the fragment condensation bstween fragmants (4) and (10) the tart- butyl ester protection was removed from the protected tripeptide (4) by treatment with 90% TFA over two hours, giving the peptide acid (11). The pentapeptide (10) was then hydroganolysed over palladium charcoal giving the amino component (12) which was subsequently coupled to the peptide acid (11). Coupling of the fragments was investigated using the Dpp mixed anhydride method and the Castro reagent [(banzotriaeol-l-yl)oxy]tris(dimethylamino)phosphonium hexafluorophosphate,²² the results being summarised in Table 1.

The octapeptide (2) was also synthesised by a fragment condensation approach utilising the sub-fragments (10) and (6). The protected tripeptide (6) was hydrogenolysed to give the amino component (13) which was than condensed with the peptide acid (14) derived from the protected pentapeptide (10) by treatment with 90% trifluoroacstic acid. In addition to the two *coupling* reagents mentioned above, di-isopropyl-carbodiimide in the presence of N-hydroxybenzotriazole was also evaluated, the results again being summarised in Table 1.

The Castro reagent (BOP) was investigated, as it was thought that this would provide a synthesis of optically pure peptide, as the rapid reaction is thought to lead to the formation of the hydroxybenzotriazolyl ester which may preclude racemisation. Alternative evidence, 17 however, suggests that considerable racemisation can occur when hydroxybenzotriazolyl active esters are formed by the conventional reaction between DCCI and hydroxybenzotriazole particularly vhen long reaction times are permitted.

The results shown in Table 1 indicate that in the preparation of octapeptides (1) and (2) the use of the BOP reagent in DMF led to a heterogeneous product as indicated by hplc and the observed optical rotations were lower than those obtained by other methods. Surprisingly, the diphenyl phosphinic mixed anhydride procedure employing activation at -20°C for one hour led to a single peak on hplc with a higher optical rotation than had been achieved by other methods. The octapeptide (2), which was obtained by the di-isopropyl-carbodiimide procedure, showed at least three poorly resolved peaks on hplc. These results are similar to earlier findings²³

TABLE 1 Coupling methods used for the octapeptides (1) and (2).

 $^+$ Activation temperature was 0^0 C except where indicated a -20[°];

a one hour activation period was used for all couplings.

which indicate that condensation of the peptide fragments containing leucine at the C -terminus gives rise to racemisation when DCCI/hydroxybenztriazole is used for activation.

Generally, low yields were encountered in the fragment coupling as indicated in Table 1, although somewhat higher yields were obtained using the BOP reagent. The various difficulties encountered using the fragment condensation strategy led us to consider a total stepwise synthesis of the octapeptide (1).

The octapeptide (1) was prepared from the pentapeptide (10) shown in Scheme 3 by further sequential elongation with Z-(He)Lau-OH, Z-Val-OH and Z-(Me)Lau-OH to give the hexa-. hepta- and octapeptides (IS), (16) and (1) respectively. The diphenylphosphinic mixed anhydride procedure^{15,24} was used for chain extension, and a 1.2-fold excess of the acylating component was employed in order to maximise the yield. The efficiency of these couplings is exemplified by the yields obtained, (96. 75 and 94% respectively) and by the fact that single peaks were observed on hplc of each peptide. No indication of the presence of diastereoisomers was evident from the hplc or 1 H nmr, although it is accepted that low levels of racemisation could conceivably pass undetected.

The high yields and overall level of purity suggest that diphenylphosphinyl chloride is particularly efficient as a coupling reagent when the point of coupling involves sterically hindered and/or N-methylated residues. In addition, as has been found previously, 15,24 that no "wrong-way" opening of the mixed anhydride was observed; thus this mixed anhydride procedure is particularly advantageous for the assembly of extensively N-methylated peptide such as cyclosporin A. The procedure thus permits stepwise assembly of large fragments of cyclosporin A and provides an alternative to the fragment condensation approach.

Acknowledgements

We acknowledge the generous financial support provided by British Technology Group (BTG) which enabled us to carry out this work.

EXPERIMENTAL

The general chromatographic and spectroscopic methods were those described previously, 15 except that:- hplc was carried out using a Spectra Physics SP8700 fitted with a 25 x, 0.46 cm ODS Chromopak C18 column eluting with CH₂CN/H₂O gradients at a flow rate of 1.5 cm^{-/}min. Gel filtration was carried out using Sephadex* LH2O or Sephadex GlO eluting with DMP and water respectively. Fractions were monitored by uv absorbance at 280 nm (Unicord III) or by optical rotation (Thorn-Bendix 143D polarimeter). In some cases column eluant was also monitored by hplc at 230 nm.

<u>Amino acid derivatives.</u>

Z-Ala-OH, Z-Val-OH, Z-Abu-OH, Z-Sar-OH, Z-Thr-OH and Z-Lau-OH were synthesised from the

and Z-(Me)Val-OH were synthesised from the corresponding ne method of Cheung and Benoiton.²¹ 2-(Me)Thr(Bu }-OH, oil, [a]²
NO_c (C, H, N); Z-(Me)Leu-OH, m.p., 74 - 75^OC_{pr}[α]² - 25.8^OC, (H, N); <u>Z-(Me)Val-OH</u>, m.p., 69 - 70°C, [ɑ]^{o - 1}86.4°, (c 5.5,

Z-(Me)Val-OBu' and

-OBu^r were each dissolved in methanol and hydrogenolysed in the presence of 10% Pd/C catalyst for twenty-four hours. The catalyst was removed by filtration and the filtrate evaporated to give an oil which was subsequently purified by fractional
distillation (0.5 mmHg).

 $\frac{H-(Me)Va1-OBu}{H, 11.30; N, 7.49}$, colourless oil, (89% yield), b.p., 44^OC. Calculated for C₁₀H₂₁NO₄; C, 63.96; H, 11.30; N, 7.49. Found: C, 63.51; H, 11.10; N, 7.2%; δ. (220 MHz, CDCl₃), 0.94 (6H. d, β-CH₃), 1.47 (9H, s, OBu^c), 1.87 (1H, m, β-CH), 2.39 (3H, s, N-CH₃), 2.77 (1H, d, α-C-H), and
7.44 (1H, br.d., N_IH), m/g 187 (M⁺, EI). (IH, br.d., N_rH), m<u>/z</u> 187 (M', EI).
<u>H-(Me)Leu-OBut</u> Colourless oil, (717

 $\frac{H - (Me) Leu - Obu^2}{H}$ Colourless oil, (71% yield), b.p., 48°C. Calculated for C₁₁H₂₄NO₀: C,
65.51; H, 11.52; N, 6.95. Found: C, 65.82; H, 11.35; N, 7.01%; S., (220 MHz, CDCI₃), 0.91 (6H, m, γ-CH₃), 1.41 (2H, m, β-CH₂-), 1.48 (9H, s, OBu), 1.73 (1H, m; γ-CH), 2.37 (1H, t, α-C-<u>H</u>),
and 7.32 (1H, br.d., NH), m/z 201 (M , EI).

General procedures

General tart-butvlation of C-terminal carboxvl group and hvdroxy side chain of amino acids

The N-benzyloxycarbonyl-N-methyl amino acid (0.21 mol) was dissolved in dichloromethane
cm³), in a pressure vessel containing concentrated H SO (5 am³). The butulation wessel (300 cm⁻), in a pressure vessel containing concentrated H_2SO_4 (5 cm⁻). The butylation vessel was cooled to -40 to -60°C, and condensed isobutylene gas (450 cm²) was added with gentle stirring. five days. The vessel was then sealed and the reaction mixture stirred at room temperature for The vessel was then cooled to -10°C , the seal removed and excess isobutylene allowed to evaporate. The carbonate (25g/l50 cm golution was allowed to reach 0 C and then brought to pH 9 with sodium eCatı The solvent was removed by evaporation and the oily residue was dissolved in ethyl acefate, washed successively with 1 M NaHCO₃, and water then dried over anhydrous Na₂SO₄. Solvent evaporation gave the <u>tert</u>-butylated product which was generally
purified by silica gel chromatography using one of a variety of solvent systems. Coupling using the diphenylphosphinic mixed anhydride method

A solution of the N-protected amino acid (1.1 equiv.) and NMM (2 equiv.) in THF (2.4 mM/ cm⁻) was cooled to -20°C and DppCl (1.1 equiv.) added. The reaction mixture was stirred for ten to twenty qinutes at -2O'C and a pre-cooled solution of the amino component (1 equiv.) in THF (2.4 mM/cm⁻) added. The suspension was stirred for one to two hours at -20°C and at ambient temperature for 24 - 48 hours. The solvent was evaporated and the residue taken up into ethyl acetate, washed successively with brine, lM KHSO into ethyl acetate, washed successively with brine, lM KHSO₄, lM NaHCO₃ and water. The organic
phase was dried over Na₂SO₄ and evaporated to give a residual oil or solid containing trace **fiin** phase was dried over Na₂SO₄ and evaporated to give a residual oil or solid containing trace
amounts of diphenylphosphinic acid. Products were generally purified by silica gel chromatography using a variety of solvent systems.

Coupling using the DICI/HOBt method

Diisopropylcarbodiimide (DICI) (1 equiv.) in DMF or THF (1 mM/2.5 cm³) was added to a mixture of the N-protected amino acid or peptide (1 equiv.), the amino component (1 equiv.), HOBt (1 equiv.) and NMM (1 equiv.). in DlIp or THP at 0 C. The mixture was stirred at 0 C for one hour and at room temperature for 12 - 20 hours. The precipitate was removed by filtration and the filtrate washed successively with **IM KHSO₄, NaHCO₃ and water, then dried over Na₂SO₄ and** evaporated to give an oil or solid which was generally purified by silica gel chromatography using various solvent systems or by chromatography,on Sephadex LH2O eluting with DMF. Coupling using Castro reagent [Bt-OP(NMe_n), PF_c 1

Benzotriazol-l-yl-oxy-tris-(dimethylamino)-phosphonium hexafluorphosphate (1.2 equiv.) was added to a solution of N-protected amino acid or peptide (1 equiv.), the amino component (1 equiv.), NMM (2 - 3 equiv.) in DMF or THF (1 mM/5 cm⁻) at O^oC, and the reaction mixture stirred at room temperature for 24 - 72 hours. The residue, after solvent evaporation was purified by silica gel chromatography or on a Sephadex LH20 column eluting with DHP.

Scheme 1

 $Z-Va1-(Me)$ Leu-OBu^t (3)

THP (oQ cm⁻) and NMM (18.7 cm⁻, $\frac{\gamma-1-\gamma}{\gamma-1-\gamma-1}$ (36.84g, 0.14 mM) in THP (60 cm⁻) was activated with DppCl (34.9g, 0.14 mM) in 0.14 mM) and coupled with $H-(Me)$ Leu-OBu (24.3g, 0.12 mM) in THF (60 cm) according to the general procedure. The mixture was stirred at -2O'C for one hour and room temperature for twenty-four hours. The solvent was evaporated and the residue worked up as described in the general method, final purification on silica (l/2) gave the title compound (51.2g. Ba) eluting with BtOAc/DCM 91%), m.p., 54 - 56°C, $[\alpha]_D^{\sim}$ - 67° (c 1.0, CH₃OH). Calculated for $C_{2,4}H_{12}N_{2}O_{5}$: C, 66.33; H, 8.81; N, 6.45. (220 MHz, $CDCL_qf$, 0.91 (6H, , 66.33; H, 8.81; N, 6.45. Found: C, 6б.36; H, 8.98; N, 6.46%; δ.
m, γ–CH₃, (Me)Leu, 1.02 (6H, m, β–CH₃, Val), 1.45 (9H, s, OBu^t), (220 MHz, CDC1₃), σ.91 (6H, m, γ-CH₃, (Me)Leu, 1.02 (6H, m, β-CH₃, Val), 1.45 (9H, s, OBu⁻);
1.51 - 1.70 (2H, m, β-CH₂, (Me)Leu), 1.91 - 2.10 (2H, m, β-CH, Val, γ-CH, (Me)Leu), 3.05 (3H, s, β–CH, Val, γ–CH, (Me)Leu), 3.05 (3H, s, N-CH₃), 4.51 - 4.61 (1H, m, α-CH, Val), 5.08 (2H, s, Ph<u>CH₂), 5.21 - 5.30 (1H, m, α-CH(Me)Leu)</u>, 5.63 (IH, d, N-H Val) and 7.37 (5H, s, ArH), m/z 434 (M', DCI), R_r 12.2 min. Z-(Me)Leu-Val-(Me)Leu-OBu (4)

a) Hydrogenolysis

<u>Z-Val-(Me)Leu-OBu"</u> (3) (38g, 85 mM) in methanol (150 cm⁻) was hydrogenolysed over 10% Pd/C (5g) at atmospheric pressure for twenty-four hours. The catalyst was removed by filtration and the solvent evaporated to give a white foam (22.5g, 93%), [α] $_{\alpha}^{\infty}$ (220 MHz, CDCl₃), 0.88 - 1.09 (12H, m, γ -CH₃ (Me)Leu, β -CH₃ Val), - 25.3°С, (с 1.0, СН₂ОН); б_и (220 MHz, CDC1₃), 0.88 - 1.09 (12H, m, γ-CH₃ (Me)Leu, β-CH₃ Val), 1.45 (9H, s, OBu⁶), 1.63 =
1.92 (3H, m, β-CH₂, γ-CH (Me)Leu), 2.47 (1H, m, β-CH Va1), 2.97 (3H, s, N-CH₂), 3.80 (1H, m, 1.92 (3H, m, β-CH₂, γ-CH (Me)Leu), 2.47 (1H, m, β-CH Val), 2.97 (3H, s, N-CH₃), 3.80 (1H, m,
α-C-H Val), 5.38 (1H, t, α-C-H (Me)Leu), and 7.29 (2H, br., N<u>H₃), m/z</u> 300 (M⁺, DCI). b) Coupling a-C-H (Me)Leu), and 7.29 (2H, br., NH₂), m<u>/z</u> 300 (M ,)

n THF (30 cm³) was activated using <u>D</u>ppCl (19.9 g, 84 mM) in THK (30 cm) and NMM (9.36 cm , 84 mM) and coupled to <u>H-Val-(Me)Leu-OBu (</u>21g, 70 mM) in THF (20
cm) according to the general coupling procedure, being stirred at -20°C for one hour and at room temperature for twenty-four hours. The residue, after solvent evaporation was worked-up as described in the general method and purified by silica gel chromatography eluting with $CH_2Cl_2/EtOAc$ (3:1,), (33.2g, 85‰), [αJ_n Evaporation of the appropriate fractions gave (4) as a colourless oil, (33.2g, 85%), $[\alpha]_{1}^{--}$ - 93.4 (c 1.0, CH₃OH). Calculated for C₃₁H₅₁N₃O_c: C, 66.26; H, 9.15; N, 7.48. Found: C, 66.53; H, 9.25; N, 7.59%; δ_u (220 MHz, CDCl₃), 0.69 - 1.12 (18H, m, γ-CH₃-7.48. Found: C, 66.53; H, 9.25; N, 7.59%; δ_H (220 MHz, CDCI₃), 0.69 - 1.12 (18H, m, γ-CH₃-
(Me)Leu, β-CH₃ Val), 1.33 (9H, s, OBu¹), 1.64 - 1.91 (6H, m, β-CH₂, γ-CH (Me)Leu), 2.05 (1H, m, (Me)Leu, β-CH₃ Val), 1.33 (9H, s, OBu⁻), 1.64 - 1.91 (6H, m, β-CH₂, γ-CH (Me)Leu), 2.O5 (1H, m,
β-C-H (Me)Val), 2.87 (3H, s, N-CH₃), 3.03 (3H, s, N-CH₃), 4.71 - 4.79 (1H, m, α-CH Val), 5.15 -
5.25 (4H, m, PhCH₂

<u>Z-Abu-Sar-OBu⁻</u> (5*)* Pivaloyl chloride₃(4.2 cm⁻, Fivaloyl chloride $_3$ (4.2 cm⁻, 33.8 mM) was added dropwise to a solution of $\frac{Z-\text{Abu}-\text{OH}}{2}$ (8.0g, 33.8 mM) in THP (85 cm⁻) and NMM (3.7 cm⁻, 33.8 mM) at -18^oC, and a total of 20 minutes allowed

for the formation of the mixed anhydride. A pre-cooled solution of $\underline{H-Sar-OBu}^t$ (2.9g, 20 mM) in THF (85 cm³) was then added, and the reaction mixture stirred at room temperature for twenty-two hours. The solvent was evaporated and the residue dissolved in ethyl acetate. The organic nous. The source was vasted successively with saturated NAHCO₃, 10X H₃PO₄, NAHCO₃ and brine. Solvent evaporation after drying (Na₂SO₄), gave a residue which was subjected to silica gal chromatography eluting w

4.8 (in, m, a-on now, J.i. (a), $\frac{1}{2}$. (b)
 $\frac{1}{2}$. (Ne)Thr(Bu^L)-Abu-Sar-OBu^L (6)
 $\frac{1}{2}$. (Ne)Thr(Bu^L)-20H (1.1g, 3.5 mM) in CH₂Cl₂ (10 cm³) was activated using DppC1 (0.2g, 3.5

mM) in CH₂Cl₂ (temperature for twenty-two hours. The product was worked-up as described in the general
method, and the residue obtained was chromatographed on a silica gel column eluting with CH_2Cl_2 / method, and the residue obtained was chromatographed on a silica gel column eluting with CH₂Cl₂/
ECOAC (1:3). Exportation of the appropriate fractions gave the tille compound as a white gum
(0.9g, 697); [q] - 44.7⁰ FAB); R_t 12.0 min.

Scheme 3

 $\overline{Z-(Me)Leu-(Me)Val-OBu}^t$ (7)

 $\frac{Z-(Me)Leu-(Me)Va1-UBu}{Z-(Me)Leu-OH}$ (22.26g, 84 mM) in THF (25 cm³) was activated with NMM (9.36 cm³, 84 mM) and
DppCl (19.92g, 84 mM) in THF (25 cm³), and coupled with $\frac{H-(Me)Va1-OBu^L}{2}$ (13.09g, 70 mM) in THF (25 cm³ temperature for twenty-four hours. Solvent evaporation gave a residue which was washed in the usual way, the resulting oil being purified on a silica gel column eluting with EtOAc/DCM (1:2). usual way, the resulting oil being purified on a silica gel column eluting with EtOAc/DCM (1:2).

Evaporation of the appropriate fractions gave the title compound as a colourless oil (31.43g, 812), [al]² - 141.8⁰ (c 1

a) Hydrogenolysis

 $\frac{Z-(Me)Leu-(MeVal-OBu^t}{2}$ (7) (17.92g, 0.04 mM) in methanol (100 cm³) was hydrogenolysed in the presence of 107 Pd/C catalyst (1g) at atmospheric pressure, for twenty-four hours. The che presence of low rayo catalyst (ig) at atmospheric pressure, for twenty-four hours. The
catalyst was removed by filtration and the solvent evaporated to give the title compound as an
oily foam (11.07g, 907), [q]² - 1

b) Coupling

 $\frac{Z_5(Me)Leu-OH}{2}$ (10.04g, 36 mM) in THF (10 cm³) was activated with DppCl (8.52g, 36 mM) in THF (10 cm³) and NMM (4.08 cm³, 36 mM) and coupled with $\frac{H-(Me)Leu-(Me)/val-OBu}{2}$ (9.42g, 30 mM) in THF (10 cm³) by the gener (10 cm⁻) by the general DppC1 method. The reaction mixture was stirred at -20°C for two hours
and at room temperature for twenty-four hours and worked-up as described in the general
procedure. The residue obtained was p FAB), R_t 13.6 min.

<u>Z-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu^t</u> (9)

a) Hydrogenolysis

 $\frac{Z-(Me)Leu-(Me)Leu-(MeVali-0Bu^t)}{2-(MeLeu-(Me)Leu-(MeVali-0Bu^t)}$ (8) (11.52g, 20 mM) in methanol (100 cm³) was hydrogenolysed
in the presence of 10% Pd/C catalyst for twenty-four hours. The solution, after removal of the catalyst by filtration, was evaporated to give the title compound as a sticky foam (7.84g, 89%); c 13, CH₂OH); δ_{I} (220 MHz, CDC1₃), 0.74 - 1.14 (18H, m, B-CH₃ (state of the Oxy), 1.49 (9H, s, 0Bu³), 1.52 - 1.94 (6H, m, B-CH₃, γ -CH₃ (Me) 1.49 (9H, s, 0Bu³), 1.52 - 1.94 (6H, m, B-CH₃, γ -CH b) Coupling

 $\frac{Z-D-A1a-OH}{and NPM}$ (4.01g, 18 mM) in THF (6 cm³) was activated by DppC1 (4.25g, 18 mM) in THF (6 cm³) and NMM (1.97 cm³, 18 mM), and the resulting mixed anhydride coupled to $\frac{H-(M\omega)Leu-(M\omega)L\omega- (M\omega)Lu-OH(6)}{0.64}$ (6.6

hours and at ambient temperature for thirty hours the residue was worked-up as described in the hours and at ambient temperature for thirty hours the residue was worked-up as described in the
general DppCl procedure and purified by silica gel chromatography eluting with RtOAc/DCM -{1:2).
Solvent evaporation of the a 13.0 min.

Z-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu^t (10)

a) Hydrogenolysis

 $\frac{Z-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu^t}{2-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu^t}$ (9) (6.42g, 10 mM) in methanol (50 cm³) was hydrogenolysed in the presence of 10% Pd/C catalyst for twenty-four hours. The catalyst was removed genoves in the presence of fox Fu/C catalyst for twenty-four nours. The catalyst was removed
by filtration and the filtrate evaporated to give the title compound as a sticky white foam,
(4.76g, 93%); [α], - 126 (c 1.2, C b) Coupling

 $\frac{Z-A1a-OH}{2-Ala-OH}$ (2.40g, 10.8 mM) in THF (4 cm³) was activated using DppCl (2.55g, 10.8 mM) in THF (4 cm³) and NMM (1.2 cm³, 10.8 mM) and coupled to <u>H-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu^t (4.68g, 9</u> mM) in THF (after work-up by the general method was purified by silica gel chromatography using CH₂Cl₂/EtOAc after work-up by the general method was purified by silica gel chromatography using CH₂Cl₂/EtOAc (2:1) as eluant. Evaporation of the appropriate fractions gave the title compound as a white solid (5.18g, 882); m.p., 5

Synthesis of Z-(Me)Leu-Val-(Me)Leu-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-Obu^t (1) using the fragment condensation strategy

CONDENSATION STREET (4) $Z - (Me)Leu - QH$ (11) (0,25g, 0.44 mM) in THF (\oint_C cm³) was activated using DppCl (0.15g, 0.60 mM) in THF (5 cm³) and COUPLE (0.15g, 0.60 mM) in THF (5 cm³) and COUPLE (0.15g, 0.60 mM) in THF general Dipple coupling method. The reaction mixture was stirred at 0 to -5 U for one nour and
at room temperature for forty-two hours. The residue, after solvent evaporation was applied to
a Sephadex LH20 column eluting by stepwise assembly; $\frac{m}{\epsilon}$ 1073 (M+1, FAB); R_t 13.6 min.

Synthesis of the octapeptide Z-Ala-D-Ala-(Me)Leu-Val-(Me)Leu-(Me)Thr(Bu^t)-Abu-Sar-OBu^t (2) by the fragment condesation approach

The fragment condessation spproach $\frac{2.41a-0.04}{0.12}$ (Me) (14) (0.4g, 0.5 mH) in THF (6 cm³) and NPH (14) (0.4g, 0.5 mH) in THF (6 cm³) and NPH (11) (0.4g, 0.6 mH) in THF (6 cm³) and NPH (11) (3.4g) 0.6 mH) in T

Synthesis of octapeptide (1) by the stepwise procedure.
 $Z-(Me)Leu-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu'$ (15).

a) Hydrogenolysis

a) nyurogenolysis
 $\frac{Z-\text{Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu}^t}{2}$ (6) (29.86g, 50 mH) was dissolved in methanol (200 cm³) and hydrogenolysed in the presence of 10% Pd/C catalyst (2.7g), at atmospheric pressure for cm⁻) and hydrogenolysed in the presence of 10x Pd/C catalyst (2.98) , at atmospheric pressure for
twenty-four hours. The catalyst was removed by filtration and the filtrate evaporated to give
the title compound as a fo

br.s., NH D-Ala), 3.75 – 3.86 (1H, d x d, α-CH (Me)Val), 4.72 – 4.90 (2H, m, α-CH Ala, D-Ala),
5.56 (2H, 2t, α-CH (Me)Leu), and 7.33 – 7.89 (2H, br.m., NH₂); m<u>/z</u> 583 (M+1, FAB). b) Coupling

s activated using DppCl (14.lg. 59.7 mM) in THF (40 cm⁻) and NMM_t (5.46 cm⁻, 59.7 mM) and coupled to
H-Ala-D-Ala-(Me<u>)Leu-(Me)Leu-(Me)Val-OBu^t</u> (29g, 49.7 mM) in THF (30 cm⁻⁾, according to the general procedure. The reaction mixture was stirred at -20°C for three hours and at ambient temperature for thirty hours. The reaction was worked-up as described for the general DppCl
method, and the residue obtained was purified on a silica gel column eluting with CH₂Cl₂/EtOAc
(2:1). Solvent evaporation of t solid (36.1g, 96%); m.p., 47 - 48°C; [α]p - 173° (c 1.0, CH₃OH). Calculated for C_{A5}H₇₆N₆O₉:
C, 63.91; H, 9.10; N, 9.95; Found: C, 64.12; H, 9.32; N, 9.80%; δ_H (250 MHz, CDC1₃), 0.76 - 1.03
(24H, β-CH₃ (Me) OBu~), 1.77 (3H, m, γ-CH~ (Me)Leu), 1.58 – 1.85 (6H, m, β-CH₂ (Me)Leu), 2.17 (1H, m, β-CH
(Me)Val), 2.77 – 3.00 (9H, series of singlets, -NCH₂), 4.39 (1H, t, α-CH L-Ala), 4.71 (1H, d, α–CH (Me)Val), 4.63 – 4.87 (2H, m, α–CH D–Ala, (Me)Eeu), 5.14 (2H, s, PhCH₂), 5.39 – 5.50 (2H, m, α-CH (Me)Leu), 6.57 (1H, br.d., NH D-Ala), 6.88 (1H, br.s., NH L-Ala), and 7.38 (5H, s, ArH); <u>m/z</u> 844 (<u>M</u>', DCI), R. 13.0 min.

<u>Z-Val-(Me)Leu-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu"</u> (16) a) Hydrogenolysis

Z-(Me)Leu-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu (15) 35.8g, 42.4 mM) in methanol (200 cm) was hydrogenolysed in the presence of 10% Pd/C catalyst at one atmosphere for thirty hours. The catalyst was removed by filtration and the filtrate evaporated to give the title compound as a white foam, (28.3g, 94%), m.p., 50 - 52°C; $[\alpha]_D^2$ - 155.4° (c 1.2, CH₂OH). Calculated for $C_{\textbf{37}}$ H₇₀N₆O₆ : C, 62.39; H, 9.93; N, 11.82. CDC1,), O.76 - 1.03 (24H, m, Found: C, 61.98; H, 9.79; N, 11.88%; δ., (250 MHz, β -CH₂ (Me)Val, γ -CH₃ (Me)Leu), 1.24 - 1.36 (6H, d x d, CH₂ Ala, D-Alã), 1.45 (9H, s, ΟΒuˤ), 1.58 - 1.90 (6H, m, β-CH₃ (Me)Leu), 1.98 (3H, m, γ-CH (Me)Leu), 2.17 (1H, m, β-CH (Me)Val), 2.76 – 2.99 (12H, series of singlets, NCH₃), 4.03 (1H, br.d., -NH (Me)Ley), 4.44 - 5.50 (5H, m, a-CH), 6.26 (1H, br.d., NH Ala), 8.39 (1H, br.d., NH D-Ala); <u>m/z</u> 710 (M^{\dagger} , DCI).

b) Coupling

<u>Z-Val₅OH</u> (11.45g, 45.6 µmM) in THF (30 cm⁻) was activated using DppCl (10.79g, 45.6 mM) in THF (30 cm⁻) and NMM (5.0 cm⁻, 45.6 mMM) and the resulting mixed anhydride coupled to <u>H-(Me)Leu-</u> <u>Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu"</u> (27g, 38 mM) in THF (30 cm), as described for the general DppCl coupling procedure. The reaction mixture was stirred at -20°C for two hours and at room temperature for forty-five hours. The reaction was worked-up as described in the general procedure and the residue subjected to chromatography on silica gel eluting with $CH₂Cl₂/$ EtOAc $(2:1)$. Evaporation of the solvent gave the title compound as a white foam, $(26.9$ g, m.p., 69.5°C; [α]² - 143.1° (c 1.1 CH₃OH). Calculated for C₅₀H₈₅N₇O₁₀ : C, 63.58; H, 9.08; N,
10.38. Found: C, 63.36; H, 9.05; N, 10.48%; δ_υ (250 MHz, CDC1₃), C.71 - 1.06 (30H, φ, β-CH₃ Val, (Me)Val, γ -CH₂ (Me)Leu), 1.31 – 1.34 (6H, d⁻* d, CH₂ Ala, D-Ala), 63.36 : H. 9.05: N. 10.48%: δ. (250 MHz. $CDC123.7C31 -$ - 1.60 (3H, m, γ-CH (Me)Leu), 1.58 - 1.84 (6H, m, β-CH3 (Me)Leu, 1.44 (9H, s, OBu⁻), 1.47 1.06 (30H, φ, β-CH₃ m, β -CH (Me)Val), 2.76 - 3.01 (12H, series of singlet 1.99 (1H, m, β–CH Val), 2.14 -NCH₃), 4.36 (1H, m, α-CH **Ala**), 4.46 , m, a-CH Val), 4.68 (lH, d. a-CH (Me)Val), 4.61 - 4.84 (lH, 2, a-CH D-Ala), 5.03 - 5.13 (4H. m, PhC<u>H</u>₂, α-CH 7.01 (zfi: br.m.. (Me)Leu), 5.34 – 5.45 (2H, m, α-CH (Me)Leu), 6.58 [1H, br.d., NH D-Ala), 6.82 –
, NH L-Ala, Val), and 7.23 (5H, s, ArH); m<u>/z</u> 943 (M , DCI); R, 13.6 min. Z-(Me)Leu-Val-(Me)Leu-L-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBut (17 a) Hydrogenolysis

 $\frac{2-\text{Val}-(\text{Me})\text{Leu}-\text{Ala}-\text{MeV}\text{Leu}-(\text{Me})\text{Leu}-(\text{Me})\text{Veu}-(\text{Me})\text{Val}-\text{OBu}^t}{\text{Ca}^3}$ was hydrogenolysed in the presence of 10% Pd/C catalyst for two days. Evaporation of the cm) was hydroganolysed in the presence of 10% Pd/C catalyst for two days. Evaporation of the of the catalyst, gave the title compound as a white foam, (20.96g, 94%); - 138.9° (c 1.2, CH₂OH). Calculated for $C_{4,2}H_{2,0}N_{7}O_{9}$: C, 62.25; H, 9.84; N, 12.10. Found : C, 62.20; H, 9.65; N, 12.23%; $\delta_{\rm tr}$ (250 MHz, CDCl,), 0.71 - 1.06 (30H, γ-CH₃ (Me)Leu, β-CH₃ Val, (Me)Val), 1.23 - 1.33 (6H,"2d, CH₃-Ala, D-Ala), 1.44 (9H, s, OBu^c), l.40⁻– 1.61 (3H, m, γ-CH (Me)Leu), 1.50 – 1.84 (6H, m, β-CH₂ (Me)Leu), 1.25 – 1.99 (2H, m, β-CH Val, (Me)Val), 2.11 (2H, br., NH $_2$), 2.78 – 3.01 (12H, sefies of singlets NCH $_2$), 4.38 (1H, m, α-CH Ala), 4.45 – 5.44 (6H, α-CH (Me)Leu, (Me)Val, Val, D-Ala), 6.68 (1H, br.d., NH D-Ala), and
6.95 (1H, br.d., NH L-Ala); <u>m/z</u> 809 (<u>M</u>', DCI). b) Coupling

THF (1 8.56g, 30.7 mM) in THF (10 cm) was activated using DppCl (7.26g, 30.7 mM) in and NMM (3.37 cm , 30.7 mM) and was coupled to H–Val-(Me)Leu-Ala-D-Ala-(Me)Leu-(Me)Leu-(Me)Val-OBu^r (20.7g, 25.6 mM) in THF (50 cm⁻), according to the general coupling procedure. The reaction mixture was stirred at -20°C for two hours and at ambient temperature for forty-five hours. The solvent was evaporated and the reaction worked-up as described in the general procedure. The residue was subjected to chromatography
on silica gel eluting with EtOAc/CH_oCl_o (l:l). Evaporation of the appropriate fractions gave on silica gel eluting with EtOAc/CH₂Cl₂ (1:1). Evaporation of the appropriate fractions gave
the title compound as a white solid, (23g, 94%); m.p., 75°C; [α], 1 - 133.1° (c 1.2, CH₂OH). the title compound as a white solid, (23g, 94%); m.p., 75°C; [α] - 133.1° (c 1.2, CH₃OH).
Calculated for C₅₇H₀₉N₉O₁₁ : C, 63.87; H, 9.22; N, 10.48; Found: C, 63.74; H, 9.23; N, 10.33%; Calculated for C₅₇H₉₈N₈O₁₁ : C, 63.87; H, 9.22; N, 10.48; Found: C, 63.74; H, 9.23; N, 10.33%;
6₁₁ (250 MHz, CDC1₃, O.76 - 1.04 (36H, m, β-CH₃ Val, (Me)Val, γ-CH₃ (Me)Leu), 1.31 - 1.34 (6H, d δ_H (250 MHz, CDCI, 30 CF) 1.04 (36H, m_e β-CH₃ Val, (Me)Val, γ-CH₃ (Me)Leu), 1.31 - 1.34 (6H, d
x⁻d, CH₃ Ala, D-Ala), 1.44 (9H, s, OBu⁶), 1.47 - 1.60 (3H, m, γ-CH(Me)Leu), 1.61 - 1.70 (8H, m, x~d, CH₃ Ala, D–Ala), 1.44 (9H, s, OBu~), 1.47 - 1.60 (3H, m, γ–CH(Me)Leu), 1.61 - 1.70 (8H, m,
β–CH₃ (Me)Leu), 1.98 (1H, m, β–CH Val), 2.15 (1H, m, β–CH (Me)Val), 2.79 - 3.06 (15H, series of singIets, NCH₃), 4.39 - 4.49 (1H, m, α-CH Ala), 4.69 - 4.95 (3H, m, α-CH D-Ala, Val, (Me)Val), 5.03 - 5.23 (4H, m, α-CH (Me)Leu, PhCH₂), 5.29 - 5.52 (2H, m, α-CH (Me)Leu), 6.50 (1H, br.d., NH
D-<u>A</u>la), 6.51 (1H, br.d., NH Val), 6.84 (1H, br.d., NH L-Ala), and 7.28 (5H, s, ArH); m/z 1072 $(\underline{M}^+, \text{ DCI}), R_{t}$ 13.6 min.

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